

URINE-ID™

CASE REVIEW

PATIENT HISTORY

Patient is a 49-year-old female with UTI symptoms, negative culture; allergic to Cipro and Sulfa.

SYMPTOMS/DISEASE STATE

Patient reported burning, frequency, and urgency when urinating for the past several days.

WHY THE TEST WAS ORDERED

Patient's culture came back negative, but patient was exhibiting symptoms of a UTI and was adamant that was the cause of her symptoms. Provider determined the need for Vikor's Urine-ID™ since patient was not having relief of symptoms.

RESULTS

PATHOGENS DETECTED

Escherichia coli	1 x 10 ⁷ copies/uL	90.827%
Enterobacter aerogenes	1 x 10 ⁶ copies/uL	9.083%
Streptococcus agalactiae	1 x 10 ⁴ copies/uL	0.091%

RESISTANCE GENES DETECTED & POTENTIAL MED CLASS AFFECTED

tetM

Tetracycline



OUTCOME

Cephalexin 500mg PO BID for 7 days. Both the patient and provider were pleased Vikor Urine-ID™ was able to successfully identify the pathogen of the patient's symptoms and provide PharmD guidance for appropriate treatment. Patient reported that she was relieved of her symptoms after following the recommended treatment regimen.



VIKORSCIENTIFIC

KORPATH

22 WestEdge Street 8th Floor
Charleston, SC 29403
P: (854) 429-1069 • F: (833) 247-4091
www.vikorscientific.com



Patient Name	Date of Birth	Gender	Race
	XX-XX-1972	F	UNDISCLOSED

Facility Information

Ordering Provider:

Facility:

Facility Phone:

Facility Fax:

Specimen Information

ACC:

Collection Date: 10-08-2021

Received Date: 10-09-2021

Notes:

Report Date:

Sample Type:

Laboratory Results

PATHOGENS DETECTED

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RESISTANCE GENES DETECTED & POTENTIAL MED CLASS AFFECTED

tetM	Tetracycline	
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ABXAssist™

Pharmacy Guidance Provided by:



Electronically approved on 10-10-2021 by: Bradley Pate

Email: pharmconsult@vikorscientific.com • Phone: 1(855) 742-7635, 1(855) PharmD5

MEDICATION REVIEW

Drug Allergies: CIPRO, SULFA

Notes from Ordering Physician:

Notes from Pharmacist:

FIRST LINE

Medication	Route	Dose
		Macrobid 100mg PO BID x 5-7 days – cystitis only
nitrofurantoin	oral	Considerations: To cover E. coli and Enterobacter. Not effective in Pyelonephritis or prostatitis. Use caution and adjust dose CrCl < 60ml/min. Contraindicated CrCl < 30ml/min. Avoid use in G6PD deficiency.
OR		
fosfomycin	oral	3g every 48-72h x 1-3 doses
		Considerations: To cover E. coli and Enterobacter. Do not use in Pyelonephritis.

SECOND LINE

		500mg PO BID x 5-7 days (uncomplicated); 500mg PO BID x 10 days (complicated)
cephalexin	oral	Considerations: To cover E. coli. Does NOT cover Enterobacter. Can utilize Cefdinir if desired. Adjust dose in renal insufficiency.

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OR
 ceftriaxone intramuscular 500mg – 1gram q24h x one to three doses

Considerations: To cover E. coli. Does NOT cover Enterobacter. Can be used as monotherapy or in conjunction with other regimens.

ALTERNATIVE

gentamicin intramuscular 5mg/kg q24h IM/IV

Considerations: To cover E. coli and Enterobacter. In renal compromised pt, use Conventional Gentamicin dose: 1mg-2.5mg/kg/dose q 8-12h IV/IM. BBW for ototoxicity and nephrotoxicity.

OR
 piperacillin / tazobactam intravenous 3.375- 4.5gm IV q6h

Considerations: To cover E. coli and Enterobacter. Can utilize Cefepime, Avycaz, Meropenem, or Ertapenem if desired. CrCl 20 to 40 mL/minute: Administer 2.25 g every 6 hours; CrCl <20 mL/minute: Administer 2.25 g every 8 hours; Can be given as continuous infusion.

Methodology	The infectious disease and antibiotic resistance detection panels are tested utilizing Real-time PCR technology to detect the presence of genes associated with pathogens and antibiotic resistance via amplification of genomic DNA. Amplification and detection are performed using the Applied Biosystems™ QuantStudio™ 12K Flex Real-time PCR system, which includes the QuantStudio™ 12k Software v1.3 and Thermo Fisher Scientific TaqMan™ assays. The assays are preloaded onto TaqMan™ OpenArray plates.
Limitations	This test only detects microorganisms and antibiotic resistance (ABR) genes specified in the panel. ABR genes are detected in the specimen and are not specific to a detected pathogen. ABR genes may be detected in bacterial strains not tested for in the panel. The resistance genes for Ampicillin, selected Extended-Spectrum-Betalactamases, Vancomycin, Carbapenems, Sulfonamide, Trimethoprim, Aminoglycosides and the Quinolone gyrase groupings are assays customized by pooling the individual genes listed in the associated group. If listed as positive, this indicates that at least one of the genes in the group was detected and the class of medication could have potential resistance.
Disclaimer	This test was developed and its performance characteristics determined by Vikor Scientific™. It has not been cleared or approved by the FDA. The laboratory is regulated under CLIA as qualified to perform high complexity testing. This test is used for clinical purposes. It should not be regarded as investigational or for research. Pharmacy guidance and recommendations therein are not under the purview of the laboratory or agencies which accredit the laboratory. The treatment guidance listed in the report is based on infectious disease treatment references, the organisms detected, and genes known to contribute to medication resistance. Important clinical information such as comorbidities, renal function, patient weight, platelet count, microbiology results, etc. may influence the overall appropriateness of therapy. The provided guidance only takes drug allergies into account when they are provided and available to the pharmacist making the recommendation. The overall appropriateness of therapy must be determined by the physician treating the patient. The provider has all the patient information necessary to make that determination and should take the entire clinical presentation into account when making treatment decisions. Should the treating physician wish to discuss the provided guidance, the pharmacist is available for consult at the email and phone number provided.

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NEGATIVE PATHOGENS

- Acinetobacter baumannii
- Actinobaculum schaalii
- Aerococcus urinae
- Alloscardovia omnicolens
- Atopobium vaginae
- BVAB2
- Candida albicans
- Candida glabrata
- Candida krusei
- Candida parapsilosis
- Candida tropicalis
- Chlamydia trachomatis
- Citrobacter freundii
- Corynebacterium riegelii
- Corynebacterium urealyticum
- Enterobacter cloacae
- Enterococcus faecalis
- Enterococcus faecium
- Gardnerella vaginalis
- Haemophilus ducreyi
- HPV 16
- HPV 18
- HSV1
- HSV2
- Klebsiella oxytoca
- Klebsiella pneumoniae
- Mobiluncus curtisii, mulieris
- Morganella morganii
- Mycoplasma genitalium, hominis
- Neisseria gonorrhoeae
- Prevotella bivia
- Proteus mirabilis
- Proteus vulgaris
- Providencia stuartii
- Pseudomonas aeruginosa
- Serratia marcescens
- Staphylococcus aureus

NEGATIVE RESISTANCE GENES

- aac6-1b/aacA4, ant(3), aph(A6), aac6-1b-cr
- ampC, ACC, DHA, ACT/MIR
- SULL, DFRA
- VEB, blaNDM-1, OXA-1, GES, SHV, PER-1, PER-2
- VIM, KPC, IMP-7, OXA-48, OXA-23, OXA-72, OXA-40, OXA-58, NDM, blaOXA-48, IMP-16
- TEM, TEM E102K, TEM R162S, TEM G238S
- CTX-M
- ermA, ErmB
- mecA
- mcr-1
- QnrB, QnrA, Gyrase A D87N_GTT, Gyrase A S83L_TGG
- VanB, VanA1, VanA2

ANTIBIOTIC CLASS

- Aminoglycosides
- AmpC beta lactamase
- Bactrim
- Beta-lactams
- Carbapenems
- Class A Beta-lactams
- ClassA Beta-lactamases
- Macrolides
- Methicillin
- Polymyxins
- Quinolones
- Vancomycin

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NEGATIVE PATHOGENS

Staphylococcus epidermidis

Staphylococcus saprophyticus

Treponema pallidum (Syphilis)

Trichomonas vaginalis

Uncultured Megaspheara 1

Ureaplasma urealyticum